

A Biosensor of S100A4 Metastasis Factor Activation: Inhibitor Screening and Cellular Activation Dynamics[†]

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ABSTRACT: S100A4, a member of the S100 family of Ca²⁺-binding proteins, displays elevated expression in malignant human tumors compared with benign tumors, and increased expression correlates strongly with poor patient survival. S100A4 has a direct role in metastatic progression, likely due to the modulation of actomyosin cytoskeletal dynamics, which results in increased cellular motility. We developed a fluorescent biosensor (Mero-S100A4) that reports on the Ca²⁺-bound, activated form of S100A4. Direct attachment of a novel solvatochromatic reporter dye to S100A4 results in a sensor that, upon activation, undergoes a 3-fold enhancement in fluorescence, thus providing a sensitive assay for use *in vitro* and *in vivo*. In cells, localized activation of S100A4 at the cell periphery is observed during random migration and following stimulation with lysophosphatidic acid, a known activator of cell motility and proliferation. Additionally, a screen against a library of FDA-approved drugs with the biosensor identified an array of phenothiazines as inhibitors of myosin-II associated S100A4 function. These data demonstrate the utility of the new biosensor both for drug discovery and for probing the cellular dynamics controlled by the S100A4 metastasis factor.

S100A4, a member of the S100 family of proteins, is directly involved in driving the progression of metastatic disease. S100A4 protein levels are elevated in malignant forms of breast, gallbladder, pancreatic, prostate, esophageal, gastric, lung and thyroid carcinomas (1). Furthermore, nonmetastatic cancer cells engineered to overexpress S100A4 produce a high incidence of tumors with a shorter latency period and higher frequency of metastasis (2, 3). However, the most compelling evidence has emerged from animal studies using S100A4 overexpression or genetic deletion of S100A4 in models of osteosarcoma, breast and lung cancer, which conclusively demonstrate that S100A4 plays a direct role in tumor metastasis (4–7). These *in vivo* studies support an important but poorly understood role for S100A4 in mediating the metastatic process.

More recent efforts have focused on the prognostic significance of S100A4 expression and its use as a marker for metastasis and poor patient outcome (8, 9). In a study of 349 invasive human breast cancer specimens, those that stained positively for S100A4 were highly correlated with

patient death (8, 10). Additionally, S100A4 is a prognostic marker in other cancers, including prostate, bladder, non-small lung, pancreatic, esophageal-squamous, primary gastric and malignant melanomas (1). These studies not only illustrate the potential use of S100A4 as a marker but also suggest S100A4 as a target for novel therapeutic drugs.

S100 proteins are typically homodimers that contain N- and C-terminal EF-hands connected by a loop or hinge region (1). Ca²⁺-binding to the C-terminal EF-hand induces a significant change in the angle between helices 3 and 4 that flank the C-terminal Ca²⁺-binding loop, and exposes a hydrophobic cleft that constitutes a binding surface for target proteins (1). S100A4 specifically binds to the C-terminal end of the coiled-coil of myosin-IIA in a Ca²⁺-dependent manner (11). Our previous studies demonstrate that Ca²⁺-activated S100A4 promotes the monomeric, unassembled state of myosin-IIA by depolymerizing preassembled filaments and inhibiting the assembly of myosin-IIA monomers into filaments (11).

Myosin-IIA localizes to the lamellae of migrating cells (12, 13). Localization studies of S100A4 show that it is diffuse throughout the cytoplasm and enriched at sites of protrusion along the leading edge (14, 15). Recent studies demonstrate that S100A4-expressing cells become highly polarized upon chemotactic stimulation by extending enhanced forward protrusions and suppressing side protrusions (15). Furthermore, the ability of S100A4 to promote directional motility is an immediate consequence of its interaction with myosin-IIA (15). These observations suggest that S100A4 expression may promote a metastatic

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phenotype via the regulation of myosin-IIA assembly and cell motility.

Here we describe the development of a biosensor that reports on the Ca^{2+} -induced activation of S100A4. This biosensor has been used to examine the spatial activation of S100A4 in living cells. Additionally we describe the use of this biosensor as a screening tool to initiate a search for inhibitory compounds, thus providing the first step toward the development of novel S100A4-based therapies.

EXPERIMENTAL PROCEDURES

Protein Preparation. Wild-type human S100A4 was expressed and purified as described previously (11). S100A4 mutants C3R/C86S and C81S/C86S were cloned into the *NdeI/HindIII* sites of pET23a (Invitrogen, Carlsbad, CA). Cysteine mutants were expressed in BL21(DE3)* without IPTG induction and purified as described for wild-type S100A4. The S100A4 protein concentration was determined using the Bradford protein assay (Bio-Rad, Hercules, CA) and a S100A4 standard of known concentration. The concentration of the S100A4 standard was determined by amino acid analysis (Keck Biotechnology Resource Laboratory at Yale University, New Haven, CT).

Human myosin-IIA rods were purified as described previously (11). The concentration of the myosin-IIA rods was determined by absorbance at 280 nm.

A peptide, FITC-DAMNREVSSLKNKLRR-OH,¹ derived from the human myosin-IIA (residues 1908–1923), was synthesized using solid-phase peptide synthesis (Biosynthesis Inc., Lewisville, TX). A 5 mM stock solution of the FITC-MIIA^{1908–1923} was prepared in 20 mM Tris pH 7.5, 150 mM KCl, 1 mM DTT and 0.02% NaN_3 , aliquoted and stored at -80°C . The purity of the peptide (>95%) was determined using HPLC and mass spectrometry.

Fluorescent Labeling of S100A4. Cys81 on S100A4 was labeled in a reaction containing 100 μM C3R/C86S S100A4 in phosphate buffer (50 mM NaPO_4 pH 7.5, 20 mM NaCl, 0.02% NaN_3 and 0.25 mM TCEP) containing 10% acetonitrile and a 4.8-fold excess of I-SO-iodoacetamide (sodium-3-[(2Z)-5-[(iodoacetyl)amino]-2-[(2E,4Z)-4-(1,1-dioxido-3-oxo-1-benzothien-2(3H)-ylidene)but-2-enylidene]-3,3-dimethyl-2,3-dihydro-1H-indol-1-yl]propane-1-sulfonate). The I-SO-iodoacetamide was synthesized as described previously (16). The labeling was quenched by the addition of 1 μL of 100% β -mercaptoethanol (14.3 M). Free dye was removed by chromatography on G25 Sephadex gel filtration media in phosphate buffer containing 0.5 mM β -mercaptoethanol instead of TCEP. The dye to protein ratio was calculated by determining the protein concentration using the modified Lowry assay and the dye concentration by measuring the absorbance at 600 nm ($\epsilon = 138,000$). This method consistently resulted in greater than 90% labeling efficiency. The labeling was confirmed by ESI-MS, which detected a major peak with a mass of 12,189.0 Da. This is within 1 Da of the mass of an S100A4 monomer with a single dye attached (calculated mass of labeled monomer = 12,188.5 Da; mass of unlabeled monomer = 11,634.5 Da).

To prepare the Fluo-S100A4, the C81S/C86S S100A4 was labeled with fluorescein-5-maleimide (Invitrogen) on Cys3 as described above.

Characterization of Mero-S100A4. Fluorescence spectra were acquired on a PTI fluorimeter (Photon Technology International). Reactions contained 5 μM Mero-S100A4 or Fluo-S100A4 dimer in 20 mM Tris pH 7.5, 150 mM KCl, 0.02% NaN_3 , 1 mM DTT and either 0.3 mM CaCl_2 or 2 mM EGTA. For Mero-S100A4, excitation scans were acquired from 500 to 630 nm (em: 635 nm) and emission scans acquired from 610 to 700 nm (ex: 600 nm) using a 4 nm slit width. For Fluo-S100A4, fluorescence excitation scans were acquired from 400 to 510 nm (em: 516 nm) and emission scans acquired from 500 to 590 nm (ex: 494 nm) using a 2 nm slit width.

For cell lysate experiments, NIH-3T3 fibroblasts at 80–90% confluency were washed two times with TBS followed by the addition of 75–100 μL of lysis buffer (50 mM Tris pH 7.1, 20 mM NaCl, 1 mM DTT and 5 $\mu\text{g}/\mu\text{L}$ each of chymostatin, leupeptin, pepstatin A). After incubating on ice for 15 min, the cells were collected and sonicated at the lowest setting (three 4 s pulses). After sonication the salt concentration was adjusted to 150 mM NaCl and the lysates were clarified by centrifugation at 13,000 rpm in a tabletop microcentrifuge for 15 min at 4°C . The total protein concentration was determined using the Bradford Assay (BioRad). The lysate was added to 1.25 μM Mero-S100A4 dimer in lysis buffer with either no addition, 1 mM CaCl_2 or 5 mM EGTA. Fluorescence emission spectra were obtained as described above.

Ca^{2+} -Binding Measurements. Ca^{2+} -binding affinities were determined using a 5,5'-Br₂-BAPTA competition assay as described previously (17). Calcium was titrated into a solution containing 25 μM 5,5'-Br₂-BAPTA and 25 μM wild-type S100A4 or Mero-S100A4 monomer at 25°C in 20 mM Tris pH 7.5, 150 mM KCl, 1 mM DTT, 0.02% NaN_3 , and the decrease in absorbance monitored at 263 nm. Total Ca^{2+} in the protein preparation was less than 0.1 mol of Ca^{2+} /mol of protein. Using the program Caligator, averaged data from 4 independent experiments were fit to two Ca^{2+} binding sites using a stepwise macroscopic binding equation in the presence of chelator (18). Models for single-site binding or four independent binding sites with different affinities produced poor fits to the experimental data. Additionally, since the low affinity site (EF1) does not compete well with 5,5'-Br₂-BAPTA, lower limits for this value were obtained by monitoring the χ^2 value as a function of the low affinity constant. This approach was verified with experiments using S100B for which we obtained similar constants to those reported previously (19).

Fluorescence Ca^{2+} titration experiments were carried out in a buffer containing 20 mM Tris pH 7.5, 150 mM KCl, 1 mM DTT, 0.02% NaN_3 , using an EGTA/ Ca^{2+} buffering system (20). 5 μL aliquots from a 1 mM Ca^{2+} stock were added to a solution containing 10 μM Mero-S100A4 and 25 μM EGTA with a 2 min mixing interval at 25°C . The free Ca^{2+} concentration at each point was calculated using WebMaxChelator standard constants (21). The fluorescence emission at 634 nm for each titration point was plotted as a function of the free Ca^{2+} . Data from four independent experiments was fit using both the Hill equation and a

¹ Abbreviations: MIIA, myosin-IIA; FITC, fluorescein; LPA, lysophosphatidic acid; Mero, merocyanine; TFP, trifluoperazine.

stepwise macroscopic binding equation for two sites as described previously (22).

Circular Dichroism Spectroscopy. S100A4 (wild-type, C3R/C86S and Mero) was dialyzed into 20 mM Tris pH 7.5, 20 mM KCl, 0.25 mM TCEP, 0.02% NaN₃. Protein concentrations were determined by the Bradford protein assay (BioRad) for wild-type and C3R/C86S S100A4 and by the modified Lowry assay (Pierce, Rockford, IL) for the Mero-S100A4 using a S100A4 standard of known concentration. Spectra were collected using 12.5 μ M S100A4 dimer in a 125 μ L circular quartz cuvette on a Jasco J-700 circular dichroism spectrophotometer. The data shown represents the average of six scans for each protein. The raw CD data was transformed into molar ellipticity using the Jasco software package and the known concentration of protein in the sample.

Myosin-IIA Binding Assays. Sedimentation assays were performed as described previously (11), using 1.5 μ M myosin-IIA rods and 0.1–35 μ M wild-type S100A4 dimer or 0.1–18 μ M Mero-S100A4 dimer. Binding constants were determined by fitting the data to the equation $b = B_{\max}[S]/(K_d + [S])$.

Inhibition of assembly and promotion of disassembly assays comparing wild-type S100A4 and Mero-S100A4 were carried out as described by Li et al. (11). For promotion of disassembly assays performed in the presence of FDA-approved drugs, 100 μ M compound or 10% DMSO was added to the reaction.

Fluorescence anisotropy measurements were performed at 22 °C using a Fluoromax-3-spectrofluorometer (Jobin Yvon Inc.). Individual reactions (200 μ L) contained 100 nM FITC-MIIA^{1908–1923} and 0–30 μ M S100A4 dimer in 20 mM Tris pH 7.5, 150 mM KCl, 1 mM DTT, 0.02% NaN₃, and 0.5 mM CaCl₂. Anisotropy was measured with excitation at 494 nm and emission at 516 nm. Measurements were acquired at the magic angle corresponding to the angle of 55° between the vectors of polarization of the excitation and emission light using a *G* factor of 0.634 previously determined for FITC on this instrument. Data from three independent experiments was plotted using Graphpad Prism v4 (GraphPad Software, Inc., San Diego, CA), and the dissociation constant was calculated by fitting to a single site saturation binding equation allowing for a floating $Y_{\min}$ value.

Competition assays with lead compounds contained 15 μ M S100A4 dimer, 100 nM FITC-MIIA^{1908–1923}, 0.3 mM CaCl₂, and 0–600 μ M compound in 20 mM Tris pH 7.5, 150 mM KCl, 1 mM DTT, 0.02% NaN₃, and 0.5 mM CaCl₂, 2% DMF. Controls included experiments representing maximum anisotropy (15 μ M S100A4, 100 nM peptide and calcium) and minimum anisotropy (100 nM peptide and calcium). Experiments were performed in triplicate and fit to a sigmoidal dose response equation with a variable slope to obtain the EC₅₀. Using this value the inhibition constant for the compounds was obtained using the equation $K_i = [K_{\text{dP}} \cdot \text{EC}_{50}]/[S]$, where EC₅₀ is the concentration at half-maximal binding of the competing peptide determined from fitting the raw data, K_{dP} is the dissociation constant for the binding of FITC-MIIA^{1908–1923} to S100A4 and $[S]$ represents the concentration of S100A4 dimer.

Molecular Modeling of the Mero-S100A4. The three-dimensional structure of the dye was built and energy minimized using the BUILDER module from Insight II

Table 1: Dissociation Constants for Ca²⁺-Binding to S100A4

	dissociation constant (μ M)	
	EF1	EF2
wild-type S100A4	$\geq 574^a$	2.6 ± 1.0^a
Mero-S100A4	$\geq 447^a$	0.4 ± 0.3^a
	$\geq 830^b$	$\leq 0.5^b$

^a The dissociation constants were determined using competition studies of Ca²⁺ with the chromophoric chelator 5,5'-Br₂-BAPTA. ^b The dissociation constants were determined by monitoring the fluorescence of the Mero-S100A4 during titration with Ca²⁺. The data from both assays were fit to a stepwise macroscopic binding equation for two sites (22).

(Accelrys Software Inc., San Diego, CA). The merocyanine-S100A4 complexes were modeled using COOT, and the structures of the apo-S100A4 (PDB: 1M31) and the Ca²⁺-bound S100A4 (A.R.B. unpublished results) (23). The dye molecule was covalently attached to Cys81 through the α carbon of the iodoacetamide reactant. The conformation of the dye in both complexes was modeled to maximize surface complementarity and hydrophobic interfaces between the protein and the dye molecule. No energy minimization or molecular dynamics simulations were performed, or protein–dye interaction restraints derived from alternative methods were taken into account.

Small Molecule Screen. A library of nearly 400 FDA-approved drugs (see Supporting Information, Table 1) was screened in a 96-well plate format using 5 μ M Mero-S100A4 dimer, 0.3 mM CaCl₂ and 150 μ M compound in 50 mM Tris pH 7.5, 150 mM NaCl, 0.02% NaN₃, and 1 mM DTT (screening buffer). Controls included Mero-S100A4 in the absence of any added Ca²⁺ and Mero-S100A4 in the presence 0.3 mM CaCl₂ or 0.3 mM CaCl₂ and 2 mM EGTA. The fluorescence intensity was measured using a Molecular Probes SpectraMax M5 plate reader (ex, 600 nm; em, 634 nm).

IC₅₀ values were calculated from reactions containing 5 μ M Mero-S100A4, 1–300 μ M compound and 0.3 mM CaCl₂. Data were plotted using GraphPad Prism v4 and IC₅₀ values were determined using a three parameter logistic equation with a standard slope (24).

LPA Stimulation: Microscopy, Image Acquisition and Analysis. NIH-3T3 fibroblasts were maintained in DMEM supplemented with 10% fetal bovine serum and grown at 37 °C in a humidified atmosphere of 5% CO₂. Prior to microinjection, cells were starved in phenol red free DMEM containing 50 mM HEPES for 2 h. Cells were injected with a 1:2 ratio of Fluo-S100A4:Mero-S100A4 (60 μ M total protein) in phosphate buffered saline pH 7.1 containing 0.2 mM DTT using an Eppendorf semiautomatic microinjection system and a 37 °C heated stage. Following injection, cells were allowed to recover for 25 min and then either fixed in 3.7% formaldehyde (control) or stimulated with 1 μ g/mL lysophosphatidic acid (LPA) for 1 or 5 min and then fixed. Dishes were mounted with a coverslip using Pro-long Anti-Fade (Invitrogen/Molecular Probes).

Phase, fluorescein and merocyanine images were acquired using IPLab Spectrum software (Scanalytics, Inc., Rockville, MD) with a CoolSNAP HQ interline 12-bit, cooled CCD camera (Roper Scientific, Tucson, AZ) mounted on an Olympus IX70 microscope with a PlanApo 60 $\times$ 1.4NA oil immersion objective (Olympus, Melville, NY). For emission

ratio imaging, fluorescein (HQ470/40, HQ525/50) and merocyanine (HQ580/30, HQ630/40) filter sets were used with a custom manufactured dichroic mirror (Chroma Technology, Rockingham, VT). Cells were illuminated through a 0.6 neutral density filter using exposure times of 0.3–0.6 s and 2×2 binning. All images were analyzed using ImageJ v1.37 (NIH). Images were background subtracted. The fluorescein image, which has the strongest signal, was thresholded to generate a binary mask with all values within the cell equal to 1 and all values outside the cell equal to 0. The fluorescein and merocyanine images were multiplied by this binary mask to eliminate background. The merocyanine image was then divided by the fluorescein image. After ratioing, the images were registered after adjusting for proper pixel alignment to minimize artifacts. Cells were traced in phase, and the outline of the cell was overlaid onto the respective ratio image. Areas outside of the cell were set to zero. The ratio image was pseudocolored, creating a map of S100A4 activation, with blue representing low and red representing high activation.

Live Cell Imaging and Analysis. Mouse embryonic fibroblasts (MEF/3T3, Clontech Inc., Mountain View, CA) were used in live cell imaging. To prevent nonspecific binding of Mero-S100A4 to fibronectin-coated coverslips during microinjection, coverslips were treated with 4% octadecyltrichlorosilane (OTS; Sigma Aldrich, St. Louis, MO) in anhydrous chloroform, followed by 3 successive washes in clean chloroform, and allowed to air-dry. Excess OTS was removed by gentle polishing, and the coverslips were coated with 10 $\mu\text{g/mL}$ fibronectin (Sigma Aldrich). 4×10^4 cells were seeded per coverslip and allowed to adhere and spread for 2 h. Medium was exchanged with standard culture medium supplemented with 10 mM HEPES and 5% w/v fatty acid free bovine serum albumin (Sigma Aldrich), and incubated for an additional 2 h prior to microinjection. Cells were microinjected with a mixture containing a 2:1 ratio of Mero-S100A4:Fluo-S100A4, at a final concentration of 73 μM in buffer containing 10 mM KH_2PO_4 , 150 mM KCl, 0.8 mM DTT. Injected cells were allowed to recover for 30 min and imaged in Ham's F-12K medium without phenol red (Invitrogen, Carlsbad, CA), with 2% fetal bovine serum, 10 mM HEPES and 10 $\mu\text{g/mL}$ Oxy-Fluo supplemented with 5 mM DL-lactate (Oxyrase Inc., Mansfield, OH), in a heated closed chamber (20/20 Technologies, Wilmington, NC). Images were obtained using an Olympus 40 $\times$ 1.3NA UIS2 UPlan FL N DIC lens on an Olympus IX71 inverted microscope (Olympus, Center Valley, PA), a CoolsnapES CCD camera (Roper Scientific, Tucson, AZ), and Metamorph software (Molecular Devices, Downingtown, PA) using the filter sets described above. Cells were illuminated through a 22% neutral density filter using exposure times of 1200 ms and 300 ms for the ISO and FITC images respectively, and 2×2 binning. Cells were imaged at 5 min time intervals for 2.5 h.

Metamorph ver 7.1.3 (Molecular Devices) was used to perform the image processing and data analysis. All images were flat-field corrected and background subtracted. ISO and FITC images were carefully aligned automatically (25) to ascertain correct whole and subpixel registration. The ISO image, because it had the largest signal-to-noise ratio and therefore provided the best distinction between the cell and the background, was intensity thresholded to generate a binary mask with a value of zero outside the cell and a value

of one inside the cell. ISO and FITC images were multiplied by this binary mask to set areas outside the cell uniformly to zero to minimize noise and other artifacts. The masked ISO image was divided by the masked FITC image to yield a ratio, reflecting Mero-S100A4 activation throughout the cell. A linear pseudocolor lookup table was applied and the ratio values were normalized to the lower scale value. In every data set, the FITC and ISO images were inspected to verify that all portions used to create the ratio image had a high enough signal/noise ratio. This was especially important in thin parts of the cell where fluorescence was low. In time-lapse experiments, ISO and FITC bleached at different rates. The ratio was corrected for bleaching using published methods (26).

RESULTS

Development of a Biosensor That Reports on the Calcium-Induced Activation of S100A4. S100A4 undergoes a large conformational change upon binding Ca^{2+} at the C-terminal EF-hand, which is necessary for interactions with protein targets and to generate a biological effect (27–30). We took advantage of this Ca^{2+} -induced conformational change and created a fluorescent S100A4 biosensor by exploiting dyes designed for reporting protein activity in living cells. The merocyanine dyes exhibit bright fluorescence at long wavelengths, are photostable, and are sensitive to both solvent polarity and hydrogen bonding, which enables them to report changes in their protein environment that occur due to conformational changes or protein–protein interactions (Figure 1A) (16, 31, 32). Using our NMR structure of the apo-S100A4 and the structures of the Ca^{2+} -bound and Ca^{2+} -target peptide complex of S100B (30, 33, 34), a structure-based alignment suggested several potentially useful sites for dye attachment, including Cys81 and Cys86. Site-specific labeling was achieved by making single serine substitutions at either Cys81 or Cys86 and replacing Cys3 with an Arg, which is found in the rodent homologues. Cys76 is buried at the dimer interface and is resistant to modification. The merocyanine dye exhibits the greatest response at position 81 (Figure 1B) with a 3-fold Ca^{2+} -induced increase in fluorescence intensity. By contrast, target binding in the presence of Ca^{2+} (e.g., myosin-IIA) does not affect fluorescence intensity, producing only a small red shift. In the presence of myosin-IIA and EGTA, Mero-S100A4 displays the same fluorescence intensity as observed with EGTA alone (data not shown). This dye yields an unusually sensitive reagent for both *in vitro* screening and live cell studies as merocyanine dyes in hydrophobic environments have quantum yields >0.7 and extinction coefficients greater than $150,000 \text{ cm}^{-1} \text{ M}^{-1}$ (16).

The biosensor can also be used to assay S100A4 activation in cell lysates (Figure 1C). In the absence of exogenous Ca^{2+} , purified Mero-S100A4 and Mero-S100A4 in a fibroblast lysate exhibit comparably low fluorescence intensities. In the presence of Ca^{2+} -supplemented lysate Mero-S100A4 displays a similar 3-fold increase in fluorescence intensity as observed with the purified biosensor. These data indicate that other potential S100A4 targets do not affect the Ca^{2+} -induced fluorescence increase of Mero-S100A4.

The merocyanines are solvatochromatic dyes that display sensitivity to both solvent polarity and hydrogen bonding,

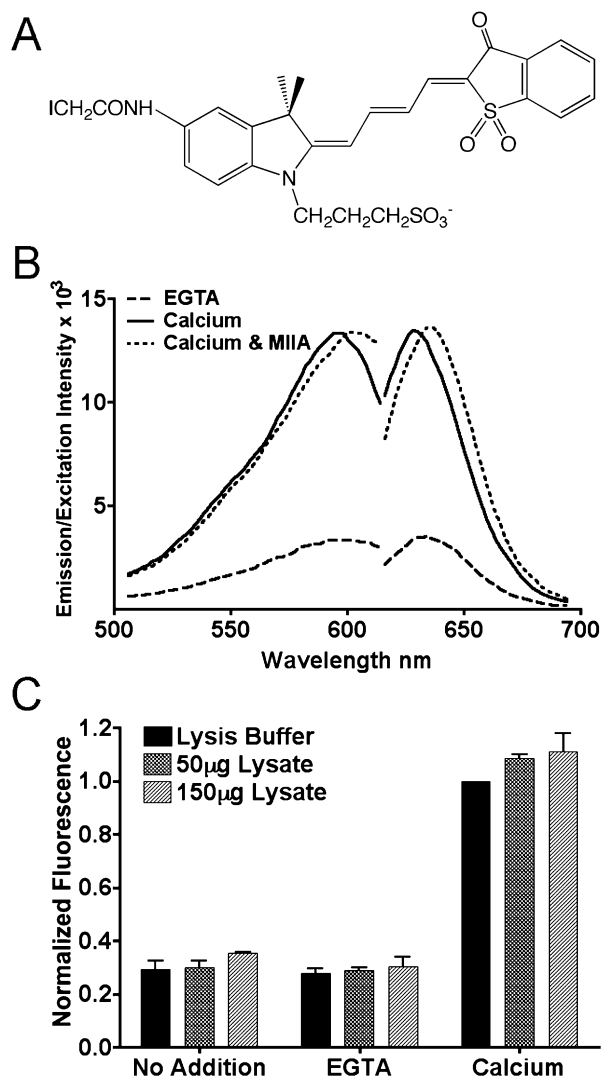


FIGURE 1: Mero-S100A4 reports activation by Ca²⁺. (A) Structure of the I-SO merocyanine dye. (B) Fluorescence excitation and emission spectra of 5 μ M Mero-S100A4 dimer. Dashed line: Mero-S100A4 in the presence of EGTA. Solid line: Mero-S100A4 in the presence of Ca²⁺. Dotted line: Mero-S100A4 in the presence of Ca²⁺ and myosin-IIA. Mero-S100A4 exhibits a 3-fold increase in fluorescence upon Ca²⁺ addition. The addition of Ca²⁺ and a 10-fold molar excess of myosin-IIA results in a slight red shift, but no additional increase in fluorescence intensity. (C) Mero-S100A4 was added to a fibroblast lysate in the presence of EGTA or calcium. The data represent the average peak intensity at 634 nm for three independent experiments and the standard deviation.

thus small changes in protein conformation can strongly influence the fluorescence properties of the dyes (16, 31, 32). Our NMR structure of the apo-S100A4 shows that helices 3 and 4 have a roughly parallel orientation with respect to one another (35). Molecular modeling of the merocyanine in the apo-S100A4 suggests that the fluorophore is solvent accessible. The structural rearrangement of helices 3 and 4 that occurs upon Ca²⁺-binding should afford the merocyanine access to the hydrophobic pocket that comprises the target binding site of S100A4 (Figure 2). This interhelical separation likely decreases the average polarity of the fluorophore's environment, resulting in an increase in the fluorescence intensity, and thus allowing the biosensor to report on the conformational rearrangements that are associated with Ca²⁺-binding in S100A4. Although the placement of the merocyanine is not expected to interfere with Ca²⁺-

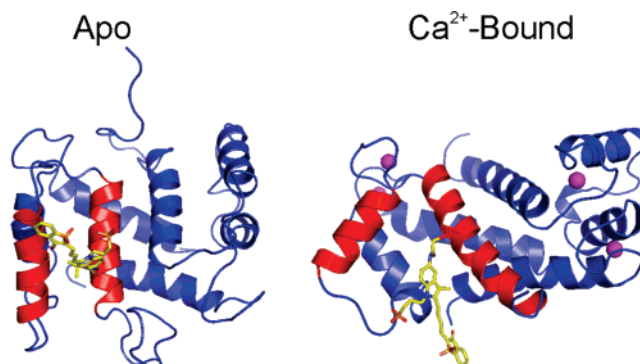


FIGURE 2: Model of Mero-S100A4. Model of the apo and Ca²⁺-bound Mero-S100A4. Helices 3 and 4 of one monomer are shown in red, and the merocyanine dye attached to Cys81 is shown in yellow. Merocyanine attached to only one monomer is shown for simplicity. The bound calcium ions are depicted as pink spheres.

binding, biochemical studies were necessary to examine potential effects of the fluorescent moiety on the Ca²⁺ and myosin-IIA binding activities of S100A4.

Ca²⁺-Binding Properties of Wild-Type and Mero-S100A4. The Ca²⁺ binding capabilities of the wild-type S100A4 and biosensor were examined in a competition assay with the chromophoric Ca²⁺ chelator 5,5'-Br₂-BAPTA (17). For the wild-type and Mero-S100A4, the dissociation constants (K_d) for the high-affinity EF-hands (EF2) are $2.6 \pm 1 \mu$ M and $0.4 \pm 0.3 \mu$ M, respectively (Table 1, Figures 3A and 3B). For the low-affinity EF-hands (EF1) K_d 's of $\geq 574 \mu$ M and $\geq 447 \mu$ M were obtained for the wild-type and Mero-S100A4 (Table 1, Figures 3A and 3B). 5,5'-Br₂-BAPTA has a relatively high affinity for Ca²⁺ (2.3 μ M), thus only limiting values could be estimated for the low affinity EF-hands (EF1) as they do not effectively compete for Ca²⁺ binding in this assay (17). These data show that Mero-S100A4 binds Ca²⁺ with slightly higher affinity at the EF2 site than wild-type S100A4 yet maintains the same affinity at the EF1 site. Previously, Ca²⁺-binding constants for S100B were reported as $\geq 350 \mu$ M and $56 \pm 9 \mu$ M for EF1 and EF2 respectively (36). Using the same 5,5'-Br₂-BAPTA assay and fitting procedure as for S100A4, we obtained comparable K_d 's of $209 \pm 60 \mu$ M and $17.5 \pm 40 \mu$ M for Ca²⁺-binding to EF1 and EF2 of S100B (Supporting Information, Figure 1).

We examined the response of Mero-S100A4 as a function of Ca²⁺ concentration in the absence of myosin-IIA (Figure 3C). A cooperative binding model provides the best fit to the titration data, and the data were fit using both the Hill equation and a stepwise two-site binding equation. Macroscopic binding constants for Ca²⁺-binding to EF1 and EF2 are $\geq 830 \mu$ M and $\leq 0.5 \mu$ M in the absence of myosin-IIA with a Hill coefficient of $n_h = 2.0$, which suggests strong positive cooperativity for Ca²⁺-binding. These are in good agreement with the values obtained from the 5,5'-Br₂-BAPTA competition assay and show the ability of the biosensor to report activation at Ca²⁺ levels attainable by the cell.

Interaction of Mero-S100A4 with Myosin-IIA. Our previous studies demonstrate that Ca²⁺-activated S100A4 promotes the monomeric, unassembled state of myosin-IIA by depolymerizing preassembled filaments and inhibiting the assembly of myosin-IIA monomers into filaments (11). Wild-type and Mero-S100A4 bind to myosin-IIA filaments with similar affinities (K_d 's of $2.1 \pm 0.3 \mu$ M and $4.3 \pm 1.1 \mu$ M,

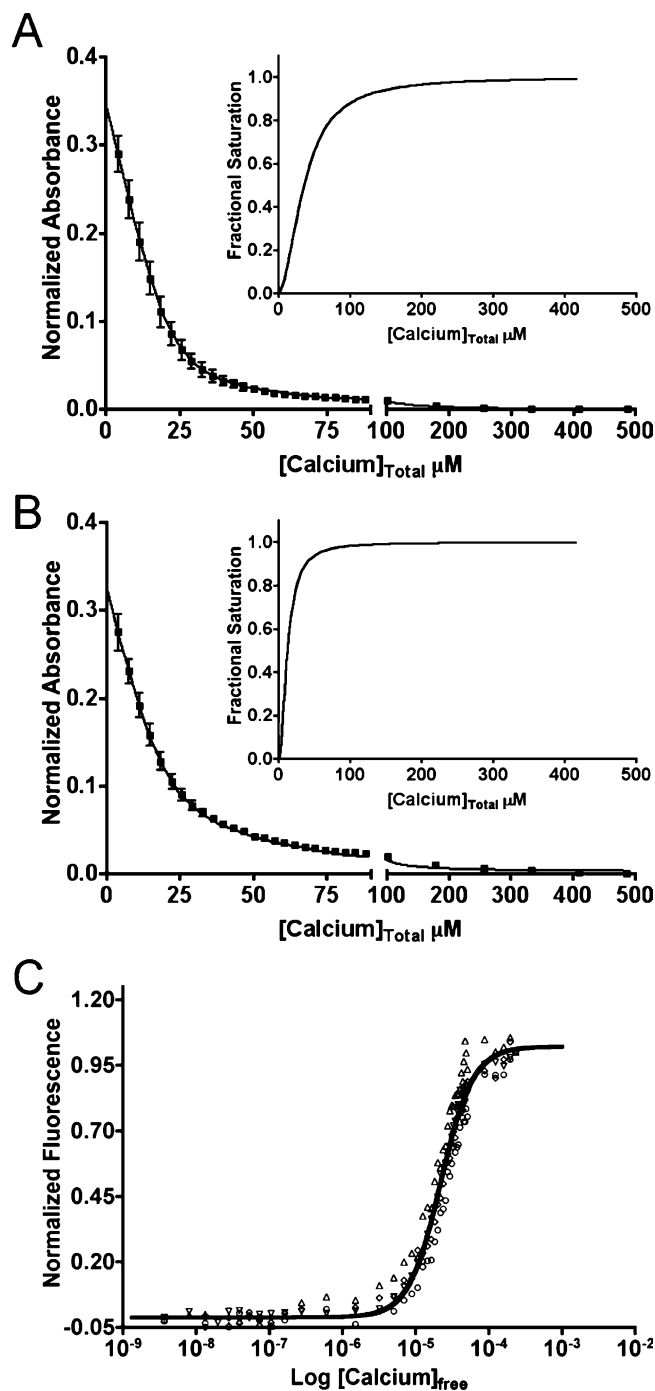


FIGURE 3: Wild-type and Mero-S100A4 bind Ca^{2+} with the same affinities. Competition assay with the chelator 5,5'-Br₂-BAPTA was used to examine the affinity of Mero-S100A4 for Ca^{2+} . The decrease in absorbance was monitored at 263 nm for a mixture containing 25 μM 5,5'-Br₂-BAPTA and either 25 μM wild-type S100A4 (A) or Mero-S100A4 (B). The data represent 6–8 titrations from 4 independent experiments. The insets show the saturation curve representation for the best fit in Caligator. (C) Ca^{2+} titration experiments were carried out using a Ca^{2+} /EGTA buffering system and 5 μM Mero-S100A4 dimer. The data represent four independent experiments.

respectively) (Supporting Information, Figure 2), indicating that dye attachment does not prevent binding to myosin-IIA. However, Mero-S100A4 does not regulate the monomer–polymer equilibrium of myosin-IIA. At a ratio of one S100A4 dimer per myosin-IIA rod, wild-type S100A4 maximally disassembles pre-existing myosin-IIA filaments and completely inhibits the assembly of myosin-IIA mono-

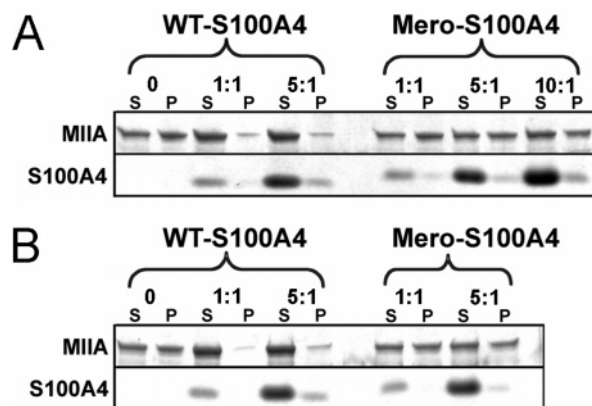


FIGURE 4: Mero-S100A4 does not regulate the monomer–polymer equilibrium of myosin-IIA filaments. (A) Promotion of myosin-IIA filament disassembly by wild-type and Mero-S100A4. At a ratio of one wild-type S100A4 dimer per myosin-IIA rod, most of the myosin-IIA rods are present in the supernatant. A ratio of five Mero-S100A4 dimers per myosin-IIA rod has no effect on the myosin-IIA filaments. (B) Inhibition of myosin-IIA filament assembly by wild-type and Mero-S100A4. At a ratio of one S100A4 dimer per myosin-IIA rod, wild-type S100A4 inhibits the assembly of myosin-IIA monomers into filaments. Mero-S100A4 does not inhibit the assembly of the myosin-IIA rods at a ratio of five Mero-S100A4 dimers per myosin-IIA rod.

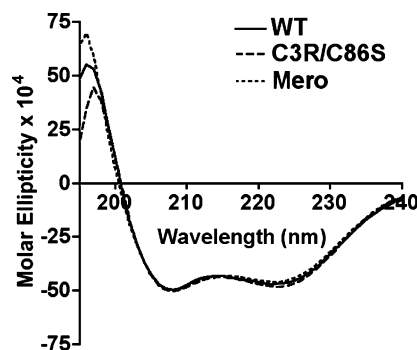


FIGURE 5: Far-UV circular dichroism spectra of wild-type, C3R/C86S and Mero-S100A4. The spectra show wild-type (solid line), C3R/C86S (dashed line) and Mero-S100A4 (dotted line) at a protein concentration of 12.5 μM dimer.

mers into filaments. At a ratio of five dimers per myosin-IIA rod, Mero-S100A4 does not destabilize pre-existing myosin-IIA filaments nor does it prevent the assembly of myosin-IIA monomers (Figure 4A and B). Circular dichroism spectroscopy indicates that the C3R/C86S mutant and Mero-S100A4 exhibit comparable secondary structures to the wild-type protein (Figure 5), therefore the inability of Mero-S100A4 to modulate the monomer–polymer equilibrium of myosin-IIA does not result from significant structural alterations in the protein.

Identification of Inhibitors of S100A4 Function. A screen against a library of FDA-approved drugs identified twelve compounds that inhibit the Ca^{2+} -induced fluorescence increase of the biosensor (Figure 6A). There was no shift in the emission maxima of Mero-S100A4 in the presence of hit compounds that would suggest inhibition caused by direct interaction with the merocyanine. Half of these compounds are members of the phenothiazine class of drugs, and include perphenazine, prochlorperazine, trifluoperazine, fluphenazine, chlorprothixene, and flupenthixol. The phenothiazines and other compounds identified in the screen display IC_{50} values in the low to mid micromolar range for inhibition of the

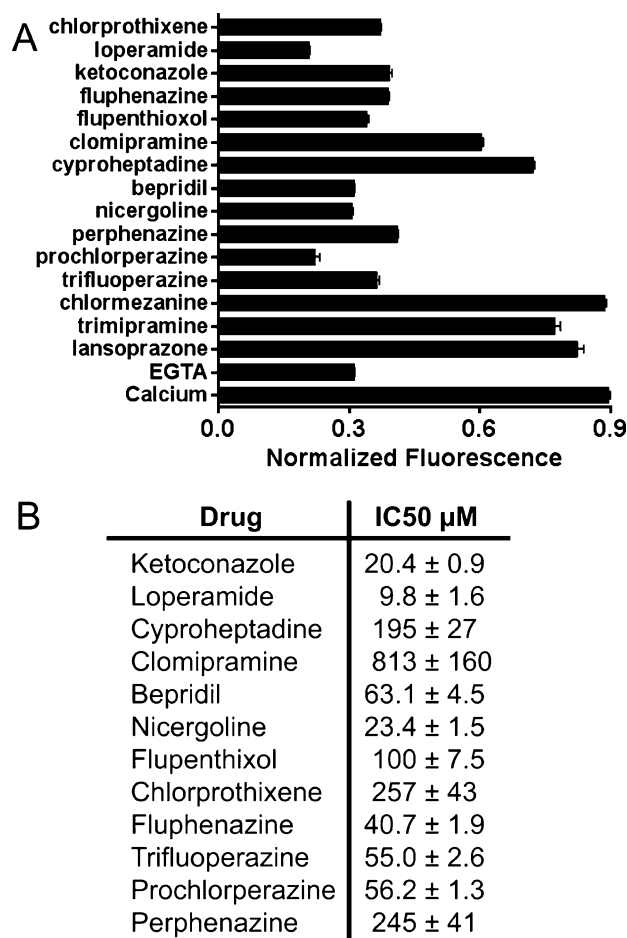


FIGURE 6: Identification of small molecule inhibitors using the Mero-S100A4. (A) Hit compounds identified in a screen of a library of FDA-approved drugs. Compounds that inhibited the Ca^{2+} -induced fluorescence increase of the Mero-S100A4 to a similar extent as EGTA were considered to be hits. (B) IC₅₀ values for inhibition of the Ca^{2+} -induced fluorescence increase of the biosensor. The data represents the best fit to a three parameter logistic equation from two independent experiments.

Ca^{2+} -induced fluorescence increase of the biosensor (Figure 6B). To eliminate the possibility that these compounds quench the fluorescence of the merocyanine dye and to evaluate their ability to inhibit the biochemical activity of S100A4, we examined their efficiency in blocking S100A4-mediated disassembly of myosin-IIA filaments (Figure 7A). Trifluoperazine completely abolishes the ability of S100A4 to depolymerize myosin-IIA filaments. Prochlorperazine, fluphenazine and bepridil also inhibit S100A4's depolymerizing activity, albeit to a lesser extent than trifluoperazine. By contrast, loperamide has no effect on the ability of S100A4 to promote myosin-IIA filament disassembly. None of the compounds affect the assembly state of myosin-IIA in the absence of S100A4. To examine whether these compounds compete with myosin-IIA for binding to S100A4, we developed a fluorescence anisotropy assay using a fluorescein-tagged myosin-IIA peptide that comprises the S100A4 binding site (FITC-MIIA^{1908–1923}). The FITC-MIIA^{1908–1923} exhibited Ca^{2+} -dependent binding to S100A4 with a K_d of $1.7 \pm 0.2 \mu\text{M}$ (Figure 7B). Titration of the S100A4-FITC-MIIA^{1908–1923} complex with lead compounds demonstrates that trifluoperazine, prochlorperazine and fluphenazine compete for myosin-IIA binding, whereas loperamide does not (Figures 7C and D). A modest increase

in the anisotropy signal was observed in assays using high concentrations of the phenothiazines as compared to controls due to the intrinsic fluorescence of the compounds at the wavelengths used to monitor the fluorescein signal. Assays with bepridil could not be performed due to insolubility at the high compound concentrations needed for effective competition. These results suggest that the phenothiazines bind to the target-binding cleft of S100A4 in a manner that inhibits myosin-IIA binding. Although loperamide inhibits the Ca^{2+} -induced fluorescence increase of the biosensor, these biochemical assays demonstrate that it does not compete with myosin-IIA for binding S100A4 nor does it promote myosin-IIA filament disassembly. These observations suggest that loperamide may interact with a region of the target binding cleft that affects the fluorescence of the merocyanine, but does not affect the interaction of S100A4 with myosin-IIA, or it may be a false positive due to direct interaction with the merocyanine.

In Vivo Imaging of the Mero-S100A4. Cell-based studies examining S100A4 activation were performed in fibroblasts since they express high levels of S100A4. Stimulation with lysophosphatidic acid (LPA), a lipid mediator that signals through a specific G-protein-coupled receptor, triggers a phospholipase C-mediated rise in intracellular Ca^{2+} . We used biosensor imaging to examine where cell surface stimulation leads to localized S100A4 activation. For *in vivo* experiments, ratiometric imaging is required to correct for variations in cell thickness, uneven illumination, and other variables that may affect the local dye intensity (37). Therefore, we created a second fluorescent analogue of S100A4 by attaching a fluorescein moiety to Cys3 (Fluo-S100A4). This analogue does not show any change in fluorescence upon Ca^{2+} or myosin-IIA binding (Supporting Information, Figure 3). To determine the level of S100A4 activation in specific subcellular compartments following LPA stimulation, the merocyanine image is divided by the corresponding fluorescein image to normalize any alterations in dye intensity that do not result from S100A4 activation. Prior to stimulation, little to no activated S100A4 is detected (Figure 8A), consistent with the low intracellular Ca^{2+} observed in quiescent cells. Upon stimulation with LPA, the biosensor reveals localized S100A4 activation at the cell periphery and in cell extensions, at 1 min poststimulation. Importantly, activation occurs concurrently with the rapid rise in intracellular Ca^{2+} that peaks within 1–2 min of LPA stimulation (38–40). By 5 min poststimulation, cells show reduced S100A4 activation, consistent with the corresponding drop in intracellular Ca^{2+} levels. These data suggest that the observed localization of S100A4 activity is due to the structural rearrangements associated with Ca^{2+} -binding, and is supported by control experiments demonstrating that cells coinjected with Mero-S100A4 and EGTA show no appreciable activation poststimulation (Figure 8B).

Time-lapse imaging of Mero-S100A4 in MEFs revealed dynamic populations of activated S100A4 present in cell edges undergoing remodeling. During random migration, S100A4 activation was observed specifically in cellular regions undergoing retraction (Figure 9). This area of elevated S100A4 activation persists even after the start of a new protrusion in the same area of the cell (Figure 9, arrowhead). These observations suggest a possible role for S100A4 in modulating edge retraction events.

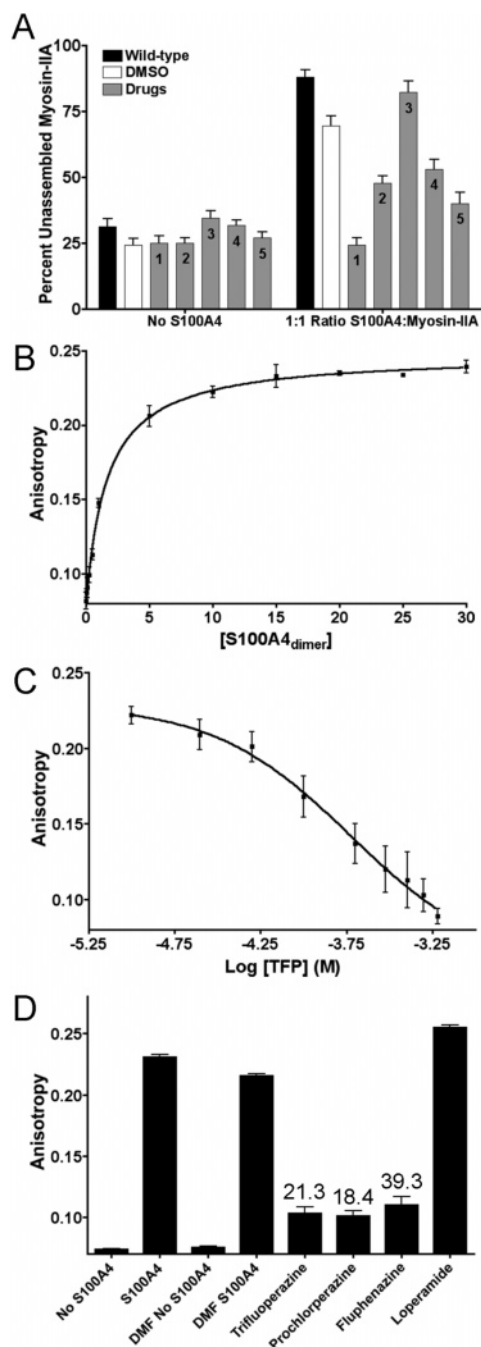


FIGURE 7: Hit compounds disrupt the interaction of S100A4 and myosin-IIA. (A) A sedimentation assay was used to examine the ability of hit compounds to attenuate the destabilization of myosin-IIA filaments by S100A4. At a molar ratio of one wild-type S100A4 dimer per myosin-IIA rod, ~90% of the myosin-IIA rods are recovered in the supernatant. Values represent the mean and the standard error of the mean from at least four independent experiments. 1, trifluoperazine; 2, prochlorperazine; 3, loperamide; 4, bepridil; and 5, fluphenazine. (B) Fluorescence anisotropy measurements of S100A4 binding to FITC-MIIA^{1908–1923}. Values represent the mean and the standard deviation from three independent experiments. A K_d of $1.7 \pm 0.2 \mu\text{M}$ was determined from the fit to a single site saturation binding curve. (C) Titration experiment with trifluoperazine showing a loss of anisotropy as TFP competes off the bound FITC-MIIA^{1908–1923}. The values represent the mean and the standard deviation from three independent experiments. The data were fit to a three parameter logistic equation to obtain the EC_{50} . (D) Bar graph representation of anisotropy competition assays with lead compounds. Values represent the mean and the standard deviation for the endpoint of the titration ($600 \mu\text{M}$ compound) from three independent experiments. The numbers above the bar indicate the K_i in units of μM .

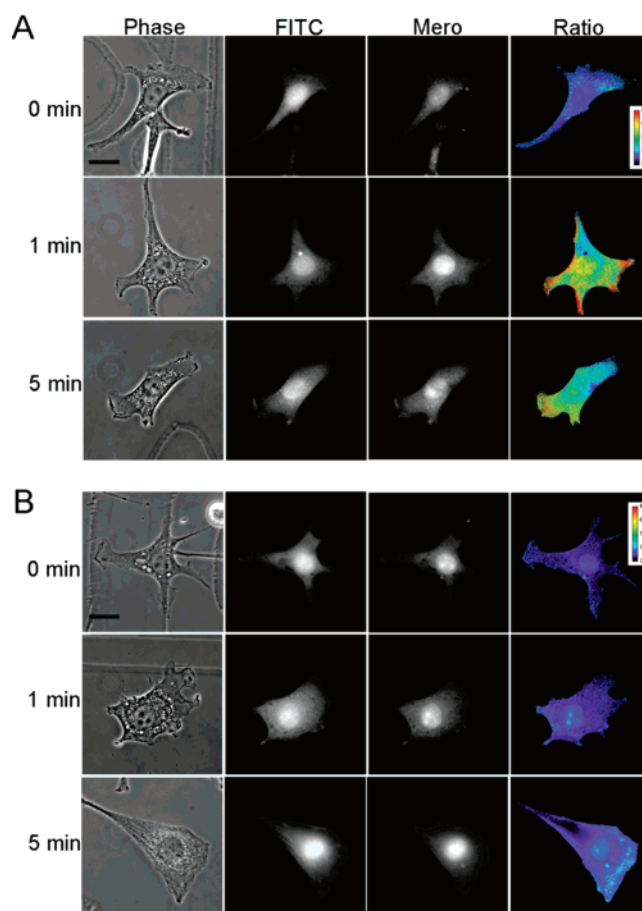


FIGURE 8: S100A4 activity in LPA-stimulated fibroblasts. (A) Cells were injected with a 1:2 ratio of Fluo-S100A4:Mero-S100A4, stimulated with LPA for the indicated times and fixed. A representative cell for each time point is shown with purple to red signifying low to high activation. At 1 min poststimulation, S100A4 activation is restricted to cell extensions with no activated S100A4 detected in the cell body. This pattern of activation was very similar in the 6 cells examined at 1 min and the 8 cells examined at 5 min. (B) Cells were injected with a 1:2 ratio of Fluo-S100A4:Mero-S100A4 in a buffer containing 1 mM EGTA, stimulated with LPA for the indicated times and fixed. A representative cell for each time point is shown with purple to red signifying low to high activation. No activation of S100A4 is detected in cells 1 min poststimulation (10 cells were examined). Scale bar = $20 \mu\text{m}$.

DISCUSSION

The direct covalent derivatization of S100A4 with a novel reporter dye has resulted in the development of the first biosensor for the S100 family of proteins. The merocyanines are solvatochromatic dyes that display sensitivity to both solvent polarity and hydrogen bonding, thus small changes in protein conformation can strongly influence the fluorescence properties of the dyes (16, 31, 32). For S100A4, conjugation with the merocyanine dye afforded a biosensor (Mero-S100A4) that reports on Ca^{2+} -induced activation and the associated structural rearrangements that result in the exposure of the hydrophobic target binding cleft. This new reagent provides a convenient and sensitive readout to screen for inhibitors *in vitro*, and to begin examining the spatial and temporal dynamics of S100A4 activation in living cells.

Biochemical characterization demonstrated that Mero-S100A4 closely mimics the wild-type protein although slight differences are observed. Ca^{2+} -binding measurements indicate that EF2 of Mero-S100A4 binds calcium ~6-fold tighter

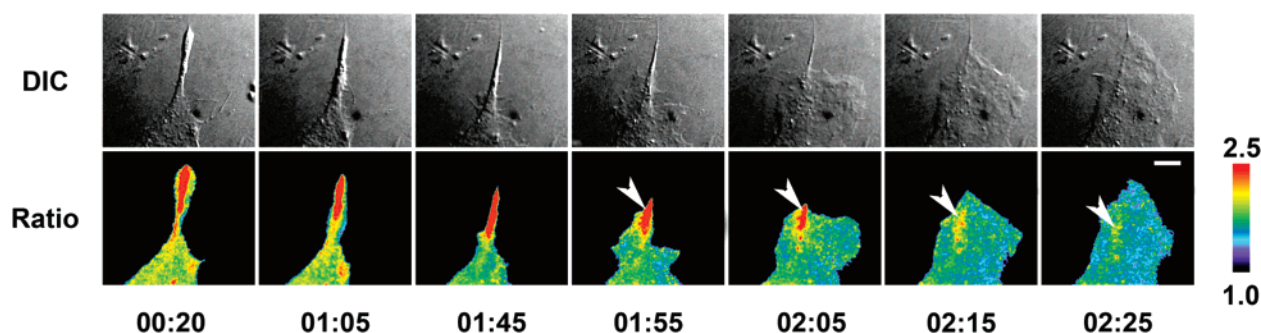


FIGURE 9: S100A4 is activated during cell retraction. The panels show different time points (hours:minutes) during random cell movement over fibronectin. Upper panels: DIC. Lower panels: ratio of Mero over FITC signal. An area of high activation is prominent during retraction. The area of elevated activation persists after the start of a new protrusion in the same area (arrowhead). Elevated Mero-S100A4 ratios were observed in cellular retractions in 5 cells examined. Scale bar = 10 μm . The ratio image was scaled by setting the lower and upper threshold levels to exclude the lower 7% and upper 3%, respectively, and a linear pseudocolor lookup table was applied where the upper threshold limit value was divided by the lower threshold limit value to show the dynamic range of ratio signal.

than the wild-type protein; however, this difference is unlikely to be significant *in vivo* due to the submicromolar (0.1–0.2 μM) concentrations of calcium in quiescent cells (41, 42). In addition, although Mero-S100A4 binds myosin-IIA, it does not disassemble filaments, thus the merocyanine moiety is associated with the target binding cleft in a manner that does not block the formation of the binary complex (S100A4–myosin-IIA). Interestingly, the phenothiazines interfere with both myosin-IIA binding and depolymerization. These observations suggest that there are distinct regions of the binding pocket that are involved in target recognition and S100A4's depolymerizing activity. More importantly, our biochemical findings suggest that the Mero-S100A4 will function as a quiescent indicator of the activated state *in vivo* since it should not perturb the monomer–polymer equilibrium of myosin-IIA.

Previous studies examining Ca^{2+} -binding to wild-type S100A4 used different methodologies from that reported here. Competition assays with the fluorescent Ca^{2+} indicator Fluo-3, which binds calcium with a 6-fold tighter affinity than 5,5'-Br₂-BAPTA, reported a K_d of 2.6 μM for two identical sites (43). This value likely reflects Ca^{2+} -binding to only the high affinity EF2 site since Fluo-3 will not compete effectively with Ca^{2+} for the low affinity sites. Moreover, this value is consistent with our measurements for the EF2 site. In studies using flow dialysis to measure Ca^{2+} -binding, K_d 's of 100 μM and 222 μM were reported for EF1 and EF2, respectively (44, 45). Several factors may account for the observed differences between these studies and those reported here. Given the difficulties associated with preparing Ca^{2+} -free protein solutions, the contaminating calcium may be sufficiently high to partially occupy the high affinity sites, resulting in titration of only the weak sites, and is consistent with the poor fitting of the data at low calcium concentrations (44). Second, increased turbidity of the protein solution was reported during titration, which further biases the measurements toward lower affinities (44). Furthermore, although our experiments were performed at physiological salt, the geometric mean constant ($\sim 36 \mu\text{M}$) is the same as that obtained by Gibbs and colleagues at low ionic strength (34–38 μM) (45). However, they reported a 5-fold lower affinity at physiological salt that, if true, would make it impossible for S100A4 to bind calcium *in vivo* even though its target interactions are calcium dependent (1, 11). This trend of reduced affinity with increasing ionic strength

could be an artifact due to the documented effects on adsorption and equilibration of the membrane during dialysis (46). Most importantly, we were able to validate our methodology by measuring Ca^{2+} -binding constants for S100B that were similar to those previously obtained.

Importantly, this biosensor has allowed for the identification of the first compounds of any kind that block S100A4 function. The use of a library of FDA-approved drugs as an initial screen to identify biological effectors of S100A4 has the following advantages: (1) the library members constitute an array of extraordinary structural diversity; (2) for nearly every library member there exists a vast number of structurally related analogues that can be examined for activity; and (3) the majority of library members are approved for human use. Therefore the identification of an effective S100A4 inhibitor has the potential of finding rapid clinical utility.

Based on the properties of the merocyanine dye and the design of the biosensor, we anticipated that our screen would identify compounds that bind within the target binding cleft of S100A4. Compounds that bind the hydrophobic cleft should prevent movement of the merocyanine into the target binding site of S100A4, resulting in an inhibition of the Ca^{2+} -induced fluorescence increase of the biosensor. Our biochemical studies of TFP binding to S100A4 are consistent with this idea. TFP binding does not inhibit Ca^{2+} -binding, rather TFP binds to the target binding cleft that is exposed due to the conformational rearrangements that occur upon Ca^{2+} -binding. Our identification of phenothiazines supports previous reports indicating interactions between S100a, S100b, S100a₀ and the phenothiazines chlorpromazine and trifluoperazine (47, 48). However, these studies only report binding of these compounds with no evidence of inhibiting or competing with target interactions. Our screen also identified several other compounds that are structurally unrelated to the phenothiazines.

Selectivity and potency are always major considerations in any drug development process. Accordingly, these issues need to be fully addressed prior to the use of phenothiazines in the treatment or prevention of metastatic disease. Phenothiazines are antipsychotics that not only are anxiolytics but also exhibit antiemetic and hypotensive attributes. These compounds may prove useful as adjuvants for cancer chemotherapy since they interfere with multidrug resistance in tumor cells (49). However, to our knowledge, these agents have not been examined as antimetastatics. At a minimum,

these studies provide proof of principle that the biosensor can be used to identify bioactive compounds that block S100A4 function.

Biosensors based on direct covalent attachment of reporter dyes to the targeted protein are used rarely in living cells, and the Mero-S100A4 represents one of the first *in vivo* applications of these new merocyanine dyes (31, 50). Given the causal role of S100A4 in promoting tumor metastasis (1), the biosensor affords a unique opportunity to examine the spatiotemporal dynamics of S100A4 activation during chemotactic motility of carcinoma cells, and will provide new information on previously inaccessible biology of a major metastasis factor. For example, pathways leading to localized activation of S100A4 can now be studied in living cells using the sensitive readout provided with this new biosensor. To test the validity of the biosensor *in vivo* we used LPA stimulation of fibroblasts as a well-characterized system to create a condition under which cells would experience a transient and globally high Ca^{2+} signal. LPA primarily promotes the release of intracellular calcium from the endoplasmic reticulum; resulting in high perinuclear calcium levels (39, 51). Our observation that S100A4 activation is highest at the cell periphery indicates that the biosensor is not merely a calcium indicator, but rather reports on the specific and localized activation of S100A4.

Previous immunofluorescence studies examining total S100A4 demonstrated localization to the leading edge and perinuclear region (14, 15). Importantly, our live cell experiments on randomly migrating cells indicate that activated S100A4 localizes to regions of the cell perimeter undergoing dynamic remodeling. In addition, the biosensor reveals that activation is transient and is associated with specific cellular activities, thus although the overall distribution of S100A4 remains relatively unchanged, activation occurs in specific subcellular locations. These experiments highlight the utility of the biosensor for deciphering the spatial and temporal dynamics of S100A4 activation, and for understanding how S100A4 regulates cell migration.

The S100 proteins share a high degree of sequence homology and a common three-dimensional fold and show similar conformational changes upon binding Ca^{2+} (35). However, most S100 family members display a high degree of target specificity, suggesting that individual S100 proteins regulate specific cellular processes (52). Moreover, alterations in S100 function are associated with a number of human diseases, including cancer, inflammatory disorders, cardiomyopathies and neurodegeneration (53–55). The successful use of the merocyanine dyes for the creation of an S100A4 biosensor provides an important proof of principle and suggests that this approach can be extended to other S100 family members.

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SUPPORTING INFORMATION AVAILABLE

Three figures, depicting Ca^{2+} -binding to S100B, binding of wild-type and Mero-S100A4 to myosin-IIA and excitation/emission spectra of Fluo-S100A4, and a table listing the compounds in the library of FDA-approved drugs. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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